

# Anti-vascular cell adhesion molecule antibody M/K-2.7–conjugated microparticles of iron oxide

VCAM-MPIO

Kam Leung, PhD<sup>1</sup>

Created: January 28, 2011; Updated: April 14, 2011.

<b>Chemical name:</b>	Anti-vascular cell adhesion molecule antibody M/K-2.7–conjugated microparticles of iron oxide	
<b>Abbreviated name:</b>	VCAM-MPIO	
<b>Synonym:</b>	M/K-2.7-MPIO	
<b>Agent category:</b>	Antibody	
<b>Target:</b>	Vascular cell adhesion molecule-1 (VCAM-1)	
<b>Target category:</b>	Antigen	
<b>Method of detection:</b>	Magnetic resonance imaging (MRI)	
<b>Source of signal/contrast:</b>	Iron oxide	
<b>Activation:</b>	No	
<b>Studies:</b>	<ul style="list-style-type: none"><li>• <i>In vitro</i></li><li>• Rodents</li></ul>	Click on <a href="#">protein</a> , <a href="#">nucleotide</a> (RefSeq), and <a href="#">gene</a> for more information about VCAM-1.

## Background

[[PubMed](#)]

Magnetic resonance imaging (MRI) maps information about tissues spatially and functionally. Protons (hydrogen nuclei) are widely used to create images because of their

---

<sup>1</sup> National Center for Biotechnology Information, NLM, NIH, Bethesda, MD; Email: MICAD@ncbi.nlm.nih.gov.

<sup>✉</sup> Corresponding author.

NLM Citation: Leung K. Anti-vascular cell adhesion molecule antibody M/K-2.7–conjugated microparticles of iron oxide. 2011 Jan 28 [Updated 2011 Apr 14]. In: Molecular Imaging and Contrast Agent Database (MICAD) [Internet]. Bethesda (MD): National Center for Biotechnology Information (US); 2004-2013.

abundance in water molecules, which comprise >80% of most soft tissues. The contrast of proton MRI images depends mainly on the nuclear density (proton spins), the relaxation times of the nuclear magnetization (T1, longitudinal; T2, transverse), the magnetic environment of the tissues, and the blood flow to the tissues. However, insufficient contrast between normal and diseased tissues requires the use of contrast agents. Most contrast agents affect the T1 and T2 relaxation times of the surrounding nuclei, mainly the protons of water. T2\* is the spin–spin relaxation time composed of variations from molecular interactions and intrinsic magnetic heterogeneities of tissues in the magnetic field (1). Cross-linked iron oxide nanoparticles and other iron oxide formulations affect T2 primarily and lead to a decreased signal. On the other hand, paramagnetic T1 agents, such as gadolinium (Gd<sup>3+</sup>) and manganese (Mn<sup>2+</sup>), accelerate T1 relaxation and lead to brighter contrast images.

Endothelial cells have important roles in inflammatory responses (2, 3). Bacterial lipopolysaccharide (LPS), virus, inflammation, and tissue injury increase tumor necrosis factor  $\alpha$  (TNF $\alpha$ ), interleukin-1 (IL-1), and other cytokine and chemokine secretion. Emigration of leukocytes from blood is dependent on their ability to roll along endothelial cell surfaces and subsequently adhere to endothelial cell surfaces. Inflammatory mediators and cytokines induce chemokine secretion from endothelial cells and other vascular cells and increase their expression of cell-surface adhesion molecules, such as intracellular adhesion molecule-1 (ICAM-1), vascular cell adhesion molecule-1 (VCAM-1), integrins, and selectins. Chemokines are chemotactic toward leukocytes and toward sites of inflammation and tissue injury. The movement of leukocytes through endothelial junctions into the extravascular space are highly orchestrated through various interactions with different adhesion molecules on endothelial cells (4).

VCAM-1 is found in very low levels on the cell surface of resting endothelial cells and other vascular cells, such as smooth muscle cells and fibroblasts (5-9). VCAM-1 binds to very late antigen-4 (VLA-4) integrin on the cell surface of leukocytes. IL-1 and TNF $\alpha$  increase expression of VCAM-1, P-selectin, and other cell adhesion molecules on the vascular endothelial cells, which leads to leukocyte adhesion to the activated endothelium. Furthermore, VCAM-1 expression is induced by oxidized low-density lipoproteins under atherogenic conditions (10). Overexpression of VCAM-1 by atherosclerotic lesions plays an important role in their progression toward vulnerable plaques, which may erode and rupture. Microparticles of iron oxide (MPIO) are composed of iron particles with a diameter of ~4.5  $\mu\text{m}$ . MPIO targeted with anti-VCAM-1 monoclonal antibody (mAb) M/K-2.7 (VCAM-MPIO) is being developed as a non-invasive agent for VCAM-1 expression in vascular endothelial cells during different stages of inflammation in atherosclerosis and renal ischemia (11).

### Related Resource Links:

- Chapters in MICAD ([VCAM-1](#))
- Gene information in NCBI ([VCAM-1](#))
- Articles in Online Mendelian Inheritance in Man (OMIM) ([VCAM-1](#))

## Synthesis

[PubMed]

The synthesis of VCAM-MPIO was described by McAteer et al. (12). The *p*-toluenesulphonyl-MPIO (diameter, 1  $\mu\text{m}$ ) was purchased commercially (Invitrogen). The M/K-2.7 mAb (33 pmol/ $10^7$  MPIO) was incubated with activated MPIO for 20 h at 37°C. IgG control MPIO (IgG-MPIO) was prepared in the same way with 33 pmol M/K-2.7 mAb. MPIO was washed and incubated with 0.1% bovine serum albumin for 4 h at 37°C to block the remaining activated sites. The number of antibodies per MPIO was not reported.

## In Vitro Studies: Testing in Cells and Tissues

[PubMed]

McAteer et al. (12) performed *in vitro* cell-binding assays with VCAM-MPIO using mouse sEND-1 endothelial cells. As determined with light microscopy, VCAM-MPIO bound to only TNF-activated sEND-1 cells ( $R^2 = 0.94$ ;  $P = 0.03$ ) in a dose-dependent manner, whereas IgG-MPIO did not. A corresponding decrease in MRI signal intensity of the MPIO was observed on 11.7-T MRI images ( $R^2 = 0.98$ ,  $P = 0.01$ ), indicating the presence of VCAM-MPIO on the cell surface. VCAM-MPIO binding was blocked by preincubation with soluble VCAM-1 and not by ICAM-1. Unconjugated MPIO was phagocytosed by mouse peritoneal macrophages and not by sEND-1 cells.

## Animal Studies

### Rodents

[PubMed]

Akhtar et al. (11) performed MRI (9.4 T) with VCAM-MPIO in a renal ischemia reperfusion injury (IRI) mouse model. The left renal pedicle was ligated for 30 min with subsequent perfusion for 16–18 h and injection of 4.5 mg iron/kg VCAM-MPIO ( $n = 5$  mice) or IgG-MPIO ( $n = 3$  mice). MRI was performed at 0, 30, 60, and 90 min after MPIO injection with automated segmentation and quantification of contrast volume. There was a rapid increase in contrast volume at 30 min and maximal contrast by 60 min in the IRI kidneys, with sequestration in the spleen and liver. IRI kidneys ( $5,991 \pm 354 \times 10^6 \mu\text{m}^3$ ) showed a 69-fold increase, but a decrease in signal in VCAM-MPIO contrast compared to the control kidneys ( $87 \pm 7 \times 10^6 \mu\text{m}^3$ ;  $P < 0.001$ ). IgG-MPIO showed minimal accumulation ( $\sim 100 \times 10^6 \mu\text{m}^3$ ) in the IRI and control kidneys. Pretreatment with anti-VCAM-1 antibody (1.32 nmol/kg, 15 min) blocked VCAM-MPIO accumulation by  $\sim 90\%$  to  $625 \pm 80 \times 10^6 \mu\text{m}^3$ . VCAM-1 expression in the IRI kidneys was 65-fold higher than in the control kidneys and in significant ( $P < 0.01$ ) correlation with VCAM-MPIO contrast volume ( $R^2 = 0.90$ ). Histological examination of the IRI kidney sections showed that

VCAM-MPIO adhered to the vessel walls of the peritubular capillaries and was not associated with local infarction and renal tubules.

## Other Non-Primate Mammals

[PubMed]

No publication is currently available.

## Non-Human Primates

[PubMed]

No publication is currently available.

## Human Studies

[PubMed]

No publication is currently available.

## References

1. Wang Y.X., Hussain S.M., Krestin G.P. *Superparamagnetic iron oxide contrast agents: physicochemical characteristics and applications in MR imaging*. Eur Radiol. 2001;11(11):2319–31. PubMed PMID: 11702180.
2. Cybulsky M.I., Gimbrone M.A. Jr. *Endothelial expression of a mononuclear leukocyte adhesion molecule during atherogenesis*. Science. 1991;251(4995):788–91. PubMed PMID: 1990440.
3. Lowe J.B. *Glycosylation in the control of selectin counter-receptor structure and function*. Immunol Rev. 2002;186:19–36. PubMed PMID: 12234359.
4. Vanderslice P., Woodside D.G. *Integrin antagonists as therapeutics for inflammatory diseases*. Expert Opin Investig Drugs. 2006;15(10):1235–55. PubMed PMID: 16989599.
5. Bochner B.S., Luscinskas F.W., Gimbrone M.A. Jr, Newman W., Sterbinsky S.A., Derse-Anthony C.P., Klunk D., Schleimer R.P. *Adhesion of human basophils, eosinophils, and neutrophils to interleukin 1-activated human vascular endothelial cells: contributions of endothelial cell adhesion molecules*. J Exp Med. 1991;173(6):1553–7. PubMed PMID: 1709678.
6. Kume N., Cybulsky M.I., Gimbrone M.A. Jr. *Lysophosphatidylcholine, a component of atherogenic lipoproteins, induces mononuclear leukocyte adhesion molecules in cultured human and rabbit arterial endothelial cells*. J Clin Invest. 1992;90(3):1138–44. PubMed PMID: 1381720.
7. Leung K.H. *Release of soluble ICAM-1 from human lung fibroblasts, aortic smooth muscle cells, dermal microvascular endothelial cells, bronchial epithelial cells, and keratinocytes*. Biochem Biophys Res Commun. 1999;260(3):734–9. PubMed PMID: 10403835.

8. Luscinskas F.W., Cybulsky M.I., Kiely J.M., Peckins C.S., Davis V.M., Gimbrone M.A. Jr. *Cytokine-activated human endothelial monolayers support enhanced neutrophil transmigration via a mechanism involving both endothelial-leukocyte adhesion molecule-1 and intercellular adhesion molecule-1*. J Immunol. 1991;146(5):1617–25. PubMed PMID: 1704400.
9. Nagel T., Resnick N., Atkinson W.J., Dewey C.F. Jr, Gimbrone M.A. Jr. *Shear stress selectively upregulates intercellular adhesion molecule-1 expression in cultured human vascular endothelial cells*. J Clin Invest. 1994;94(2):885–91. PubMed PMID: 7518844.
10. Aikawa M., Libby P. *The vulnerable atherosclerotic plaque: pathogenesis and therapeutic approach*. Cardiovasc Pathol. 2004;13(3):125–38. PubMed PMID: 15081469.
11. Akhtar, A.M., J.E. Schneider, S.J. Chapman, A. Jefferson, J.E. Digby, K. Mankia, Y. Chen, M.A. McAteer, K.J. Wood, and R.P. Choudhury, *In vivo quantification of VCAM-1 expression in renal ischemia reperfusion injury using non-invasive magnetic resonance molecular imaging*. PLoS One, 20105(9): p. e12800 (1-10).
12. McAteer M.A., Schneider J.E., Ali Z.A., Warrick N., Bursill C.A., von zur Muhlen C., Greaves D.R., Neubauer S., Channon K.M., Choudhury R.P. *Magnetic resonance imaging of endothelial adhesion molecules in mouse atherosclerosis using dual-targeted microparticles of iron oxide*. Arterioscler Thromb Vasc Biol. 2008;28(1):77–83. PubMed PMID: 17962629.