ImPyβImPyβImβ- C_3 - 18 F

[¹⁸F]PIPAM5

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Chemical name:	$ImРуβImРуβImβ-C_3-^{18}F$	
Abbreviated name:	[¹⁸ F]PIPAM5	
Synonym:		
Agent category:	Macromolecule	
Target:	DNA	
Target category:	Nucleic acid binding molecule	
Method of detection:	Positron emission tomography (PET)	
Source of signal/contrast:	18 _F	
Activation:	No	
Studies:	 In vitro Rodents	No structure is current available in PubChem.

Background

[PubMed]

Polyamides (PAM) constructed from N-methylpyrrole (Py), N-methylimidazole (Im), 3-chlorothiophene (Ct), and N-methylhydroxypyrrole (Hp) amino acids comprise a class of synthetic oligomeric ligands that bind to the minor groove of DNA (1, 2). The aromatic heterocycles in the PAM orientate antiparallel with respect to the Watson-Crick base pair (bp), which leads to a specific recognition of DNA sequences (3). The recognition process follows a series of pairing rules; i.e., an ImPy specifies for G·C, a PyPy binds both A·T and T·A, an HpPy discriminates T·A over A·T, and a CtPy prefers T·A over A·T at the N-terminus. These aromatic amino acids can be programmed to a strand with more than two residues to recognize longer DNA sequences; for example, an ImPyPy motif specifies for the five-bp sequence 5'-WGWCW-3' (W=A, T) instead of 5'-WGWWW-3' (4). More

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complicated PAM motifs can be designed by adding small molecules such as β -alanine or γ -aminobutyric acid to covalently link between two antiparallel PAM strands, yielding substantial increases in affinities and specificities. For instance, an eight-ring hairpin motif, which has a γ -aminobutyric acid (γ -turn) linker to connect the carboxylic terminus of one polyamide to the amino terminus of another, exhibits ~100-fold higher affinity for binding a six-bp DNA sequence compared to the unlinked homodimers (4). PAM are molecules that can permeate cell membranes and have been used in targeting a variety of DNA sequences in cell culture (5). The binding of PAM replaces the DNA-binding proteins and thus regulates the transcription of selected genes. The use of radiolabeled PAM aims at imaging gene regulations *in vivo*.

Fluorine-18 [¹⁸F], with a half-life of 109.7 min and low β^+ -energy (0.64 MeV), represents the ideal radionuclide for position emission tomography (PET). The ¹⁸F-produced positron is annihilated with an electron, leading to the emission of two 511-keV photons ~180° apart, which is detected coincidentally with PET. Various peptides have been successively fluorinated with multistep ¹⁸F-acylation, using ¹⁸F-labeled prosthetic groups such as amino-reactive ¹⁸F-labeling agent N-succinimidyl 4-[¹⁸F]fluorobenzoate (6). To increase labeling efficiency, the fluorination also can be conducted *via* a two-step synthetic approach in which an oxime is formed between an aminooxy group in the peptide and an ¹⁸F-labeled aldehyde such as 4-[¹⁸F]fluorobenzaldehyde (6). ImPyβImPyβImβ- C_3 -¹⁸F ([¹⁸F]PIPAM5) is an ¹⁸F-labeled PAM used for PET that is obtained with the oxime ligation approach (5). [¹⁸F]PIPAM5 contains five aromatic amino acids connected with β-alanine, which is denoted as β and is also known as a fivering β-linked motif. Its unlabeled form with an *N-N*-dimethylaminopropyl tail has been exhibit ability to upregulate the repressed gene frataxin in a cell culture mode of Friedreich's ataxia (7).

Synthesis

[PubMed]

Harki et al. reported the synthesis of [18 F]PIPAM5 (5). Initially, 4-[18 F]-fluorobenzaldehyde was obtained by nucleophilic fluorination of a trimethylammonium benzaldehyde derivative with cyclotron-produced [18 F]fluoride in the presence of 5,6-benzo-4,7,13,16,21,24-hexaoxa-1,10-diazabicyclo[8.8.8]hexacos-5-ene (Kryptofix[2.2.2]). The β-linked PAM ImPyβImPyβImβ was synthesized on a Boc-β-alanine phenylacetamidomethyl resin according to standard protocols. Then the PAM was hydroxylamine-functionalized in DMF by reaction with tert-butyl-3-aminopropoxycarbamate in the presence of benzotriazolyloxy-tris-(pyrrolidino)-phosphonium hexafluorophosphate (PyBOP) and N,N,-diisopropylethylamine. Finally, the obtained ImPyβImPyβImβ-hydroxylamine was ligated with the 4-[18 F]-fluorobenzaldehyde with aniline as a catalyst to produce [18 F]PIPAM5 at radiochemical yield of 12%. The whole synthetic procedure was completed in 100 min after the end of bombardment.

[¹⁸F]PIPAM5

In Vitro Studies: Testing in Cells and Tissues

[PubMed]

Harki et al. used the cold PAM analog [19 F]PIPAM5 to evaluate its affinity to DNA *in vitro* (5). Quantitative DNaseI footprint titrations were performed on the 5'- 32 P-polymerase chain reaction fragment from plasmid pJWP-16. In this method, equilibrium mixtures of 32 P end-labeled DNA and a range of PAM concentrations were partially digested by DNase I followed by gel electrophoresis and autoradiography. The PAM bound DNA was protected from cleavage, which produced a band gap on the gel. Quantification of the binding fraction as a function of PAM concentration was used to the apparent association constant, $3.5 \pm 2.1 \times 10^9$ M $^{-1}$ for [19 F]PIPAM5.

Animal Studies

Rodents

[PubMed]

Harki et al. examined the biodistribution of [18 F]PIPAM5 *in vivo* by PET and computed tomography (CT) (5). C57 mice were injected intravenously with [18 F]PIPAM5 at doses of 472, 218, and 193 μ Ci (17.5, 8.06 and 7.141 MBq), respectively, and PET images were collected for 2 to 3 h. At 4 min after injection, ~46% of injected [18 F]PIPAM5 was found in the liver. At 20 min after injection, 35% to 40% of injected [18 F]PIPAM5 was observed in the gastrointestinal tract and maintained a constant for the entire PET scan. No significant radioactivity was found in the brain, heart, or bone. Thus, the clearance of [18 F]PIPAM5 was primarily *via* the liver by excretion through the gallbladder and entry into small intestine; the renal clearance was <1.5%.

Other Non-Primate Mammals

[PubMed]

No publication is currently available.

Non-Human Primates

[PubMed]

No publication is currently available.

Human Studies

[PubMed]

No publication is currently available.

NIH Support

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