

(+)-8-Chloro-5-(7-benzofuranyl)-7-hydroxy-3-^[11C]methyl-2,3,4,5-tetrahydro-1H-3-benzazepine

[¹¹C]NNC 112

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Created: March 17, 2006; Updated: January 12, 2008.

Chemical name:	(+)-8-Chloro-5-(7-benzofuranyl)-7-hydroxy-3- ^{[11} C]methyl-2,3,4,5-tetrahydro-1H-3-benzazepine	
Abbreviated name:		
Synonym:	[¹¹ C]NNC 112	
Backbone:	Compound	
Target:	D ₁ dopamine receptors	
Mechanism:	Receptor binding	
Method of detection:	Positron Emission Tomography (PET)	
Source of signal:	¹¹ C	
Activation:	No	
Studies:	<ul style="list-style-type: none">• <i>In vitro</i>• Rodents• Non-primate non-rodent mammals• Non-human primates• Humans	Click on the above structure for additional information in PubChem .

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Background

[PubMed]

Dopamine, a neurotransmitter, plays an important role in the mediation of movement, cognition, and emotion (2, 3). Dopamine receptors are involved in the pathophysiology of neuropsychiatric diseases such as Parkinson's disease, Alzheimer's disease, Huntington's disease, and schizophrenia (4). Five subtypes of dopamine receptors, D₁ through D₅, have been well characterized pharmacologically and biochemically (5). These five subtypes are classified into two subfamilies: D₁-like (D₁ and D₅) and D₂-like (D₂, D₃, and D₄) dopamine receptors. D₁-like and D₂-like receptors exert synergistic as well as opposite effects at both the biochemical and overall system level. A great majority of striatal D₁ and D₂ receptors are localized postsynaptically on caudate-putamen neurons and to a lesser extent presynaptically on nigrostriatal axons.

(*R*)-(+)-8-Chloro-2,3,4,5-tetrahydro-3-[¹¹C]methyl-5-phenyl-1*H*-3-benzazepin-7-ol ([¹¹C]SCH 23390) was the first positron emission tomography (PET) radioligand for D₁ receptor studies. However, [¹¹C]SCH 23390 exhibits a low neocortex/cerebellum ratio (1.5), which makes it difficult to estimate the D₁ receptor density in the neocortex. (+)-8-Chloro-5-(7-benzofuranyl)-7-hydroxy-3-[¹¹C]methyl-2,3,4,5-tetrahydro-1*H*-3-benzazepine ([¹¹C]NNC 112) was also found to be a selective, high-affinity antagonist of D₁ receptors, but to have only a marginal effect on D₂, α₁-adrenergic, muscarinic, and histaminergic receptors and only a slight effect on 5-HT₂ receptors (6, 7). [¹¹C]NNC 112 was found to display higher specific signal/noise ratios than [¹¹C]SCH 23390. [¹¹C]NNC 112 PET has been developed to study D₁ receptor occupancy and density in the striatum and neocortex in humans.

Synthesis

[PubMed]

[¹¹C]NNC 112 was synthesized by alkylation of the desmethyl compound with [¹¹C]methyl triflate (8). Reaction in acetone with subsequent chromatographic separation gave radiochemical yields of 50-60%, with a total synthesis time of 25-30 min and a radiochemical purity >99%. The average specific activity was 111 GBq/μmol (3 Ci/μmol) at the end of synthesis.

In Vitro Studies: Testing in Cells and Tissues

[PubMed]

NNC 112 has been reported to have selective binding affinity to D₁ (striatum) and 5-HT₂ (frontal cortex) receptor sites in homogenates of rat brain membranes (6, 7). The K_i values for D₁, D₂, and 5-HT₂ were 0.18, 898, and 18 nM, respectively. The affinity for the D₂ receptor is about 4,900-fold lower than that for the D₁ receptor. NNC 112 has a 50% inhibitory concentration (IC₅₀) of 2,300 nM for the α₁-adrenergic receptor. The affinity for the 5-HT₂ receptor is about 100-fold lower than that for the D₁ receptor, suggesting that specific [¹¹C]NNC 112 binding visualized by PET represents mainly binding to D₁ receptors. Farde and Hall (9) reported that the receptor densities of D₁ to D₂ in the human putamen-caudate regions are similar and that there are more D₁ receptors than D₂ receptors in various cortical regions.

Animal Studies

Rodents

[PubMed]

Biodistribution studies in rat brain showed high accumulation of radioactivity in the striatum (2.05 ± 0.43% injected dose (ID)/g), followed by the prefrontal cortex (0.52 ± 0.10% ID/g), hippocampus (0.41 ± 0.09% ID/g), and cerebellum (0.19 ± 0.04% ID/g) at 30 min after injection of [¹¹C]NNC 112 (10). The striatum/cerebellum ratio was 10.79.

Other Non-Primate Mammals

[PubMed]

Using PET, Rosa-Neto et al. (11) directly compared the distributions of dopamine D₁ and D_{2/3} receptors in six Gottingen minipigs. The binding potentials (BPs) of [¹¹C]NNC 112 for dopamine D₁ receptors and [¹¹C]raclopride for dopamine D_{2/3} receptors were calculated in pig striatum by use of metabolite-corrected arterial inputs or with cerebellum used as the reference region. Depending on the method for quantitation, the BP for [¹¹C]NNC 112 was 1.2-5.1 in pig striatum, whereas the BP for [¹¹C]raclopride was 1.0-1.8. The reference tissue method of Logan provided the most stable estimate of BP for [¹¹C]NNC 112. Dopamine D₁ binding was relatively more abundant in the ventral-anterior striatum of the pig, whereas dopamine D_{2/3} binding was greater in the dorsal-posterior striatum. Similar comparisons were made for the BPs of [¹¹C]SCH 23390 for dopamine D₁ receptors and for [¹¹C]raclopride in the brains of six rhesus monkeys. The fraction of unchanged [¹¹C]NNC 112 in plasma was 45, 24, and 3% at 5, 10, and 60 min, respectively.

Non-Human Primates

[PubMed]

Halldin et al. (8) performed PET studies in the brains of cynomolgus monkeys and found high accumulation of radioactivity in the brain (4.2% ID) with higher accumulation in the striatum than in the cerebellum at 4 min after injection of 37 MBq (1 mCi) of [^{11}C]NNC 112. The striatum/cerebellum and neocortex/cerebellum ratios were 5.77 ± 0.31 and 2.36 ± 0.14 at 40-50 min after injection, respectively. Inhibition by the specific D₁ receptor antagonist SCH 23390 (2.5 mg/kg 21-23 min after [^{11}C]NNC 112 injection) was 80% in the striatum and neocortex at 72 min after injection. However, the 5-HT₂ antagonist ketanserin (2 mg/kg before or after [^{11}C]NNC 112 injection) showed no inhibition, indicating that binding of [^{11}C]NNC 112 to D₁ receptors in the brain is specific and reversible.

Human Studies

[PubMed]

In a PET study of D₁ receptor distribution in human brain, Halldin et al. (8) reported that [^{11}C]NNC 112 showed major localization of radioactivity in the striatum and neocortex. The striatum/cerebellum and neocortex/cerebellum ratios were 3.8 and 1.8, respectively. Transient equilibrium was achieved at 40-50 min after injection. Linear graphical analysis gave distribution volumes for the putamen, frontal cortex, and cerebellum of 15.2, 5.8, and 3.9, respectively. The fraction of unchanged [^{11}C]NNC 112 in plasma was 50% and 18% at 10 and 30 min, respectively.

Abi-Dargham et al. (12) studied 6 healthy volunteers, using kinetic and Logan graphical analysis to estimate four parameters with good reproducibility and good agreement. However, there was noise-dependent bias in the graphical but not in the kinetic analysis. Another PET study by Abi-Dargham et al. (13) in 16 patients with schizophrenia (no psychotropic medication at least 14 days before the study) and in 16 normal subjects revealed that the BP of [^{11}C]NNC 112 was significantly elevated in the dorsolateral prefrontal cortex (DLPFC) of patients with schizophrenia (1.63 ± 0.39 ml/g) compared with the controls (1.27 ± 0.44 ml/g).

Narendran et al. (14) compared regional brain BPs of [^{11}C]NNC 112 in 14 chronic ketamine users and 14 normal volunteers. There was a significant increase in [^{11}C]NNC 112 BP in the DLPFC of ketamine users (1.68 ± 0.40 ml/g) compared with controls (1.35 ± 0.35 ml/g). No significant differences were observed in other cortical, limbic, or striatal regions. Slifstein et al. (15) reported that specific binding of [^{11}C]NNC 112 in the brain cortical regions was reduced by 20-30% after administration of 2 mg of risperidone to inhibit serotonin 5-HT_{2A} receptors in 7 healthy subjects. No inhibition was observed in the striatum. These results suggested that there are ~5-10 fold selectivity of [^{11}C]NNC 112 for D₁ versus 5-HT_{2A} receptors.

Cropley et al. (16) reported on PET scans in 7 healthy volunteers after injection of ~710 MBq (19 mCi) of [¹¹C]NNC 112. Absorbed doses were found to be 32.4 μGy/MBq (0.12 rad/mCi) for the gallbladder, 22.2 μGy/MBq (0.082 rad/mCi) for the liver, 16.9 μGy/MBq (0.063 rad/mCi) for the lungs, 16.6 μGy/MBq (0.061 rad/mCi) for the kidneys, 15.7 μGy/MBq (0.058 rad/mCi) for the urinary bladder wall, 9.4 μGy/MBq (0.035 rad/mCi) for the spleen, and 6.9 μGy/MBq (0.026 rad/mCi) for the brain. The effective dose was 5.7 μSv/MBq (21.1mrem/mCi).

Supplemental Information

[Disclaimers]

Chemistry

Synthesis Protocol

Toxicology

NIH Support

Intramural research program, R01 MH59144-01, MH661710-03, K02 MH01603-0

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