

Overview of the "Matrisome"—An Inventory of Extracellular Matrix Constituents and Functions

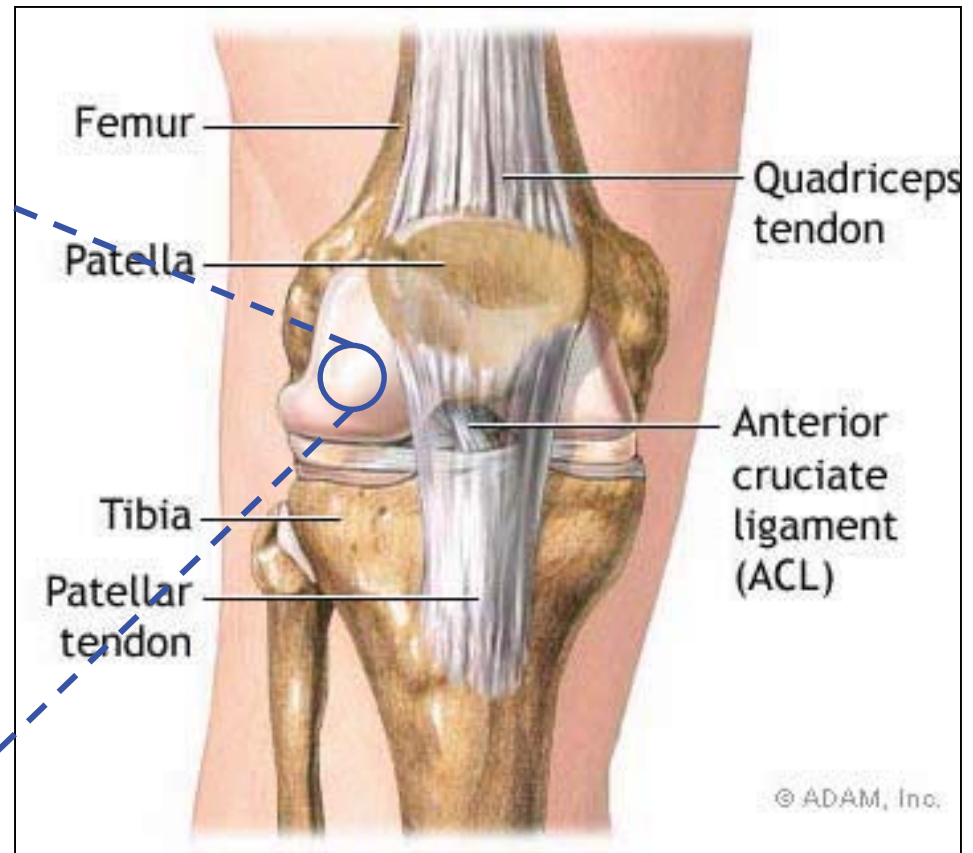
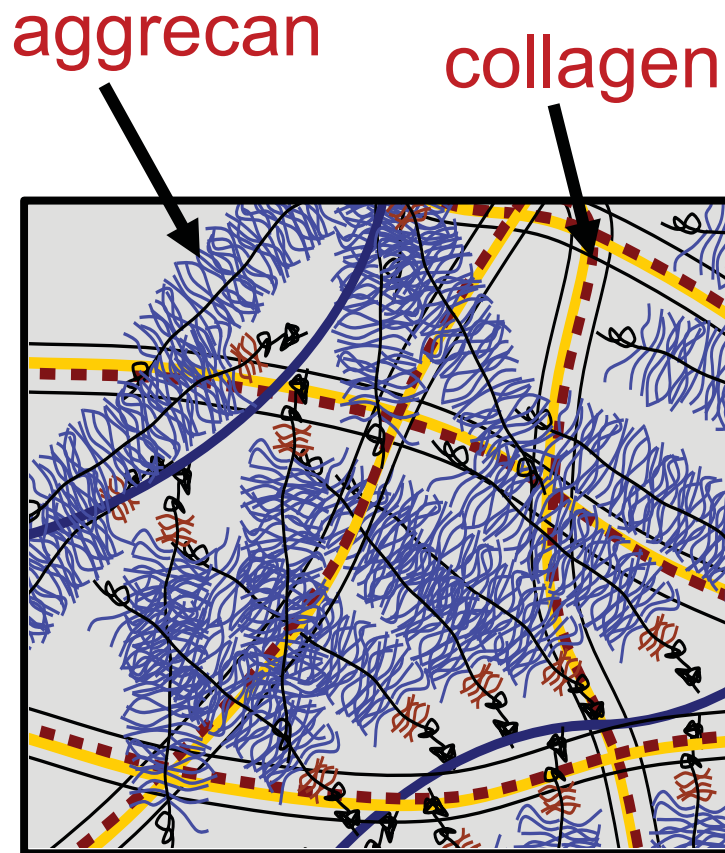
Richard O. Hynes and Alexandra Naba

Howard Hughes Medical Institute, Koch Institute for Integrative Cancer Research, Massachusetts Institute of Technology, Cambridge, Massachusetts 02139

- Completion of genome sequences for many organisms defines the complete “list” of extracellular matrix (ECM) proteins.
- In mammals: “**core matrisome**” **comprises ~300 proteins**. Also: large numbers of ECM modifying *enzymes* & ECM-binding *growth factors*
- These **ECM & ECM-associated proteins cooperate** to assemble & remodel extracellular matrices and bind to cells through receptors **so cells can survive, proliferate, differentiate, shape, and migrate**.
- **ECM proteins were the key to the transition to multicellularity, the arrangement of cells into tissues, and the elaboration of novel structures during vertebrate evolution.**

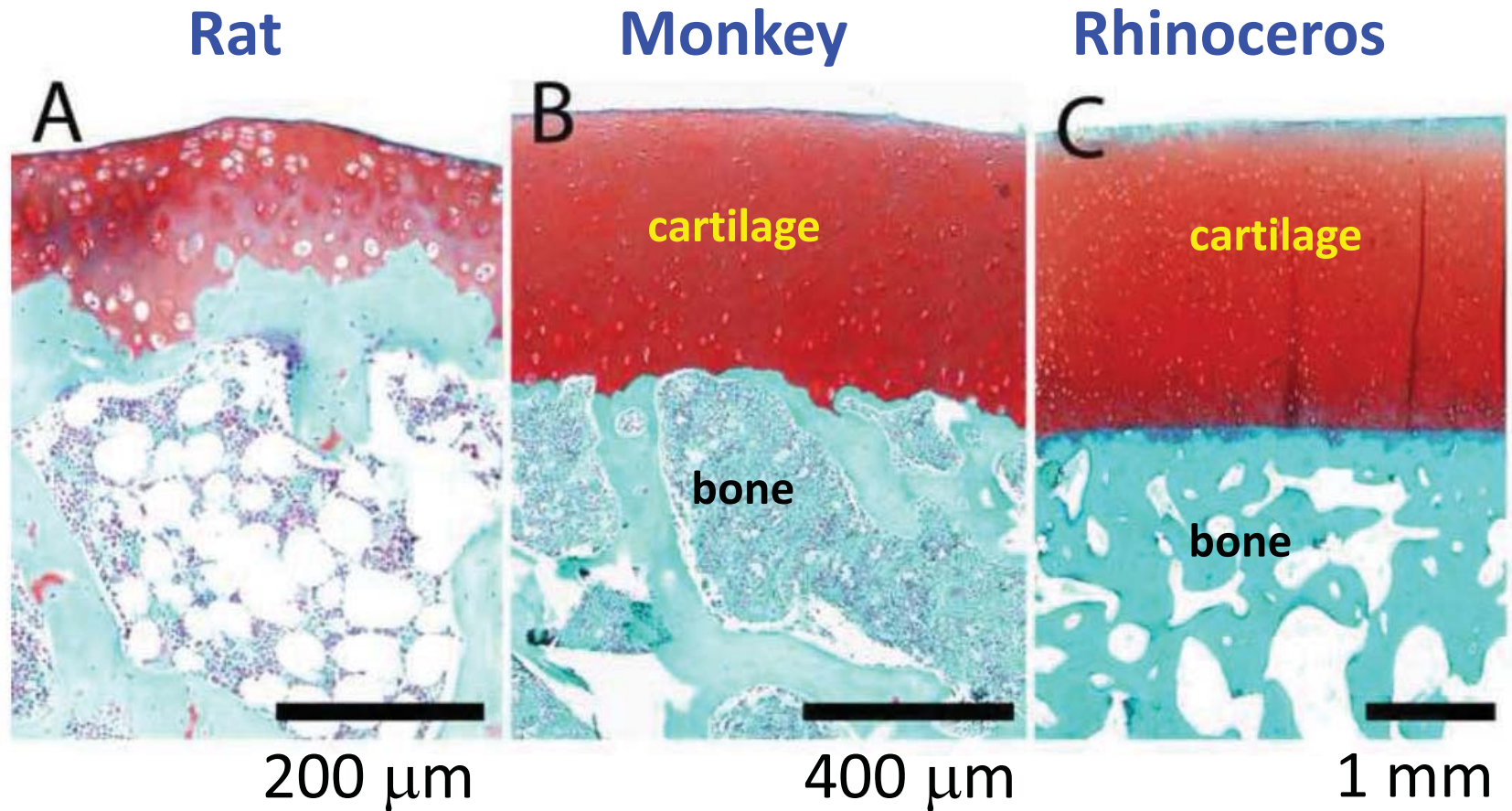
Aggrecan: Resists Compression (in Cartilage)

Collagen: Resists Tension (in Cartilage)



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"Safranin-O" (red) stains Glycosaminoglycans (of Proteoglycans)

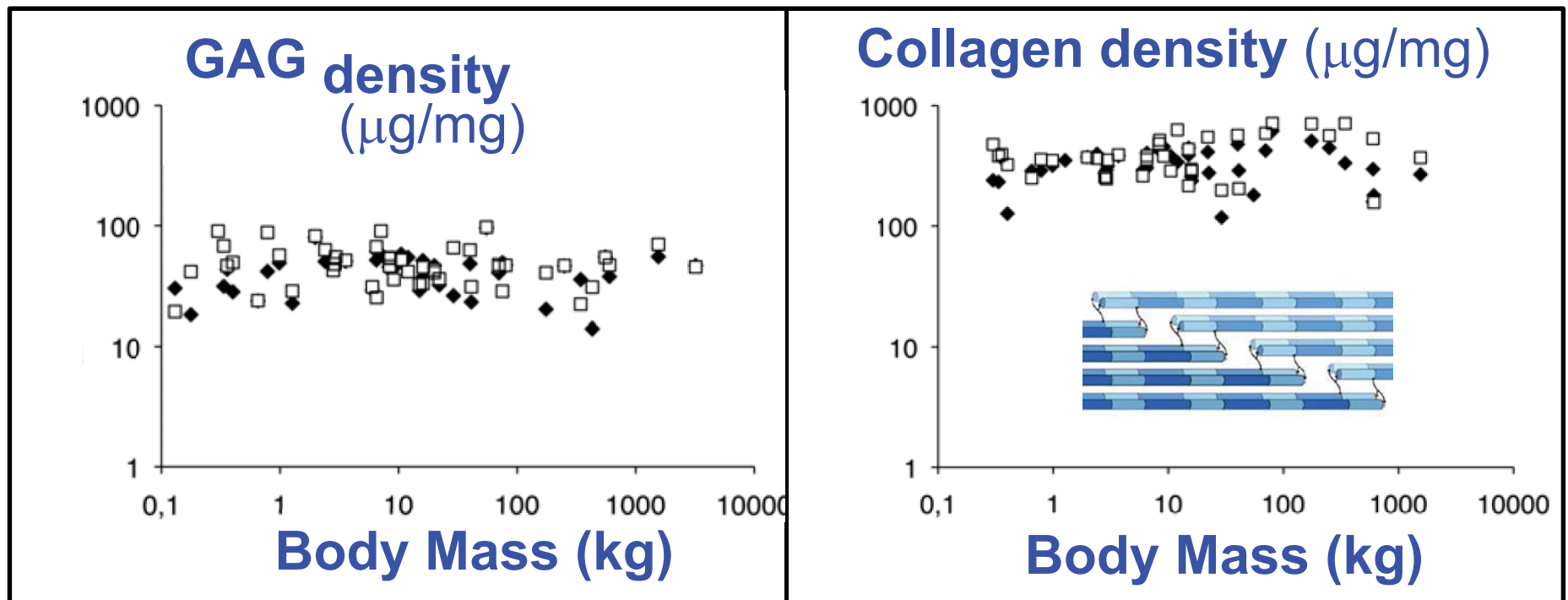
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Source: Malda, Jos, et al. "Of Mice, Men and Elephants: The Relation between Articular Cartilage Thickness and Body Mass." *D'cG'cbY 8*, no. 2 (2013): e57683.

Tensile & Shear Modulus: Collagen

Compressive Modulus: Aggrecan-GAGs

(Modulus: an "intrinsic material property"
.....independent of size, shape.....)



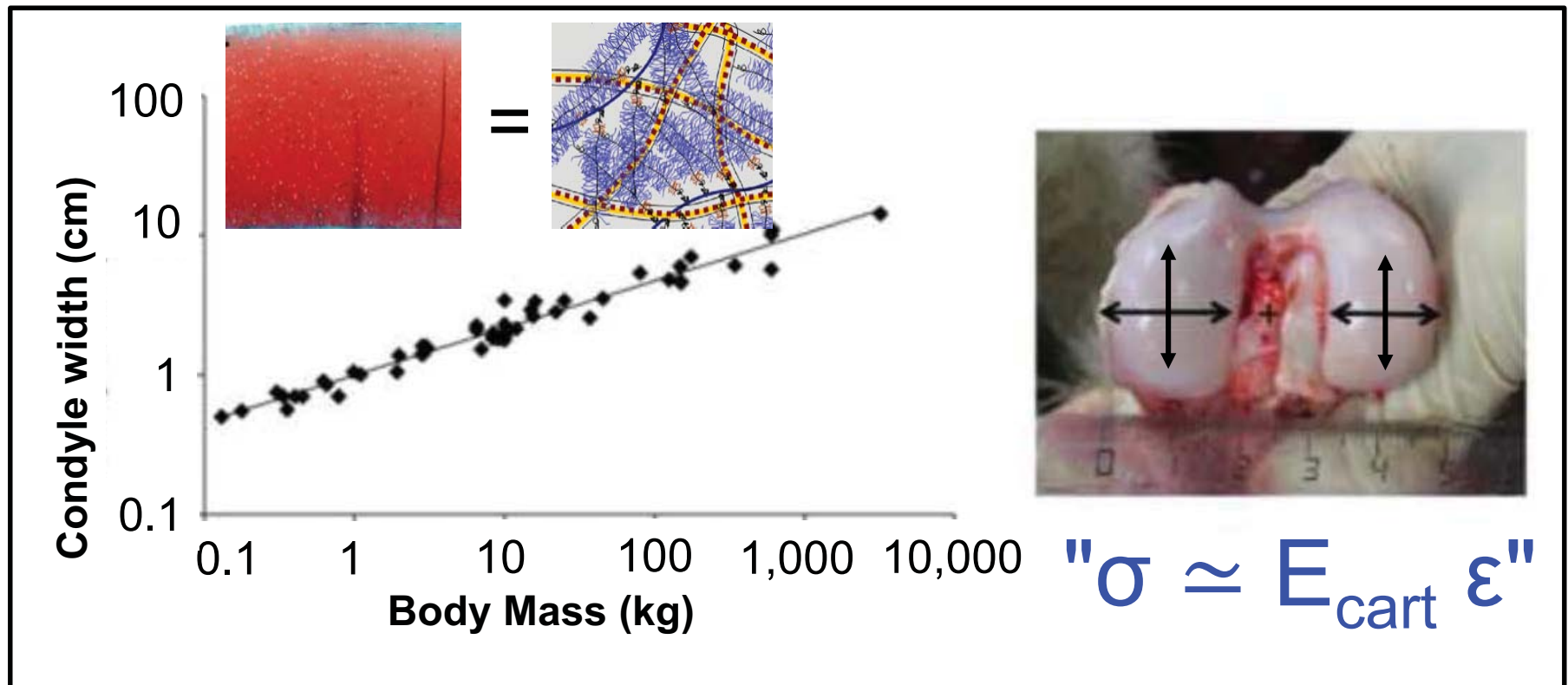
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Source: Malda, Jos, et al. "Of Mice, Men and Elephants: The Relation between Articular Cartilage Thickness and Body Mass." *D'cG'cbY* 8, no. 2 (2013): e57683.

Loading Area \propto [width]² scales as (Body Mass) ^{$\frac{2}{3}$}

Stress = (Force/Area) on Joint surface is ~ Same

\therefore cells have optimized the appropriate ECM



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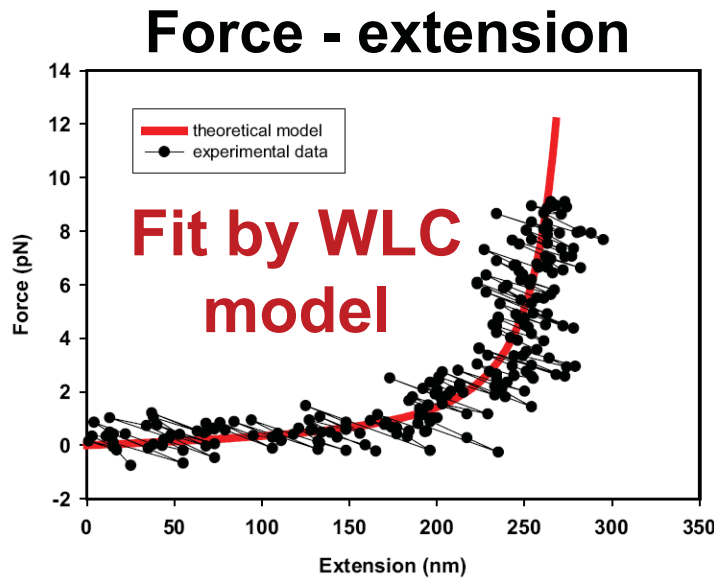
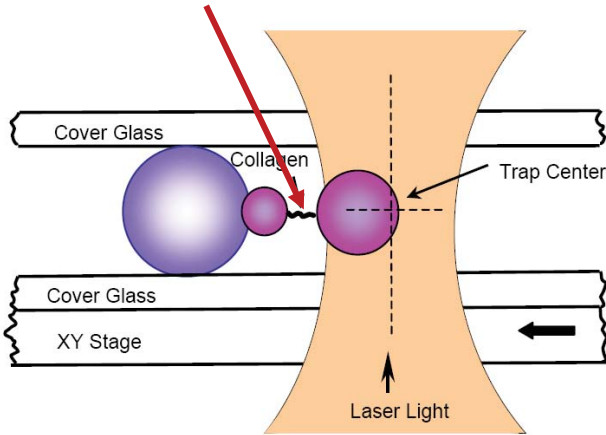
Source: Malda, Jos, et al. "Of Mice, Men and Elephants: The Relation between Articular Cartilage Thickness and Body Mass." *Discy* 8, no. 2 (2013): e57683.

Google: "elastic moduli"

Every elastic modulus can be expressed in terms of two other moduli

	bulk	Young's	Lamé #2	Shear	Poisson	Longitudinal
	$K =$	$E =$	$\lambda =$	$G =$	$\nu =$	$M = H$
(K, E)	K	E	$\frac{3K(3K-E)}{9K-E}$	$\frac{3KE}{9K-E}$	$\frac{3K-E}{6K}$	$\frac{3K(3K+E)}{9K-E}$
(K, λ)	K	$\frac{9K(K-\lambda)}{3K-\lambda}$	λ	$\frac{3(K-\lambda)}{2}$	$\frac{\lambda}{3K-\lambda}$	$3K - 2\lambda$
(K, G)	K	$\frac{9KG}{3K+G}$	$K - \frac{2G}{3}$	G	$\frac{3K-2G}{2(3K+G)}$	$K + \frac{4G}{3}$
(K, ν)	K	$3K(1 - 2\nu)$	$\frac{3K\nu}{1+\nu}$	$\frac{3K(1-2\nu)}{2(1+\nu)}$	ν	$\frac{3K(1-\nu)}{1+\nu}$
(K, M)	K	$\frac{9K(M-K)}{3K+M}$	$\frac{3K-M}{2}$	$\frac{3(M-K)}{4}$	$\frac{3K-M}{3K+M}$	M
(E, λ)	$\frac{E+3\lambda+R}{6}$	E	λ	$\frac{E-3\lambda+R}{4}$	$\frac{2\lambda}{E+\lambda+R}$	$\frac{E-\lambda+R}{2}$
(E, G)	$\frac{EG}{3(3G-E)}$	E	$\frac{G(E-2G)}{3G-E}$	G	$\frac{E}{2G} - 1$	$\frac{G(4G-E)}{3G-E}$
(E, ν)	$\frac{E}{3(1-2\nu)}$	E	$\frac{E\nu}{(1+\nu)(1-2\nu)}$	$\frac{E}{2(1+\nu)}$	ν	$\frac{E(1-\nu)}{(1+\nu)(1-2\nu)}$
(λ, G)	$\lambda + \frac{2G}{3}$	$\frac{G(3\lambda+2G)}{\lambda+G}$	λ	G	$\frac{\lambda}{2(\lambda+G)}$	$\lambda + 2G$

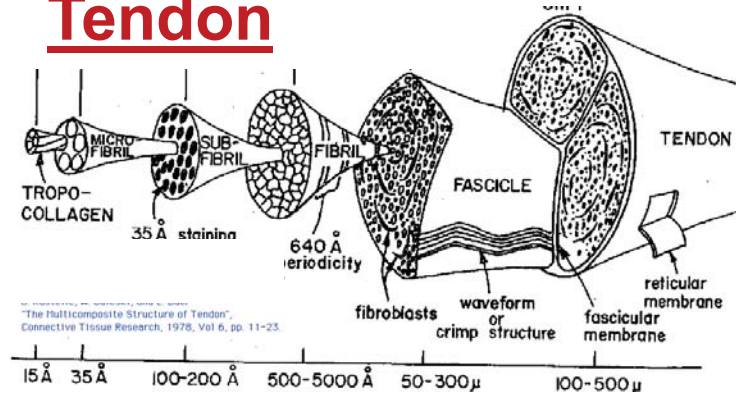
Pro-collagen molecule



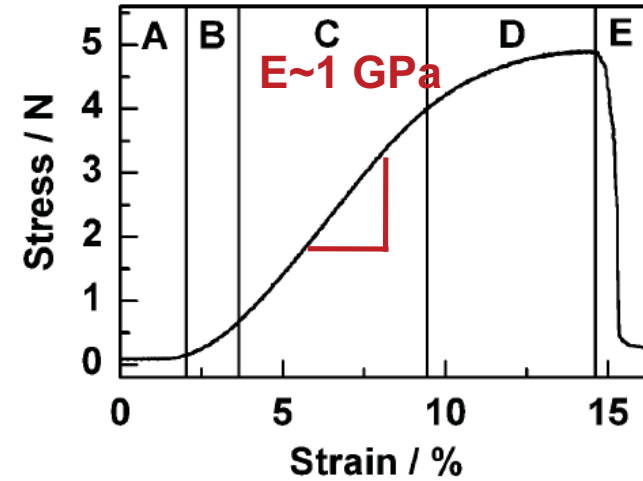
(Sun+, J Biomechanics, 2004)

Courtesy of Elsevier, Inc., <http://www.sciencedirect.com>. Used with permission. Source: Sun, Yu-Long, et al. "Stretching Type II Collagen with Optical Tweezers." *Journal of Biomechanics* 37, no. 11 (2004): 1665-9.

Tendon



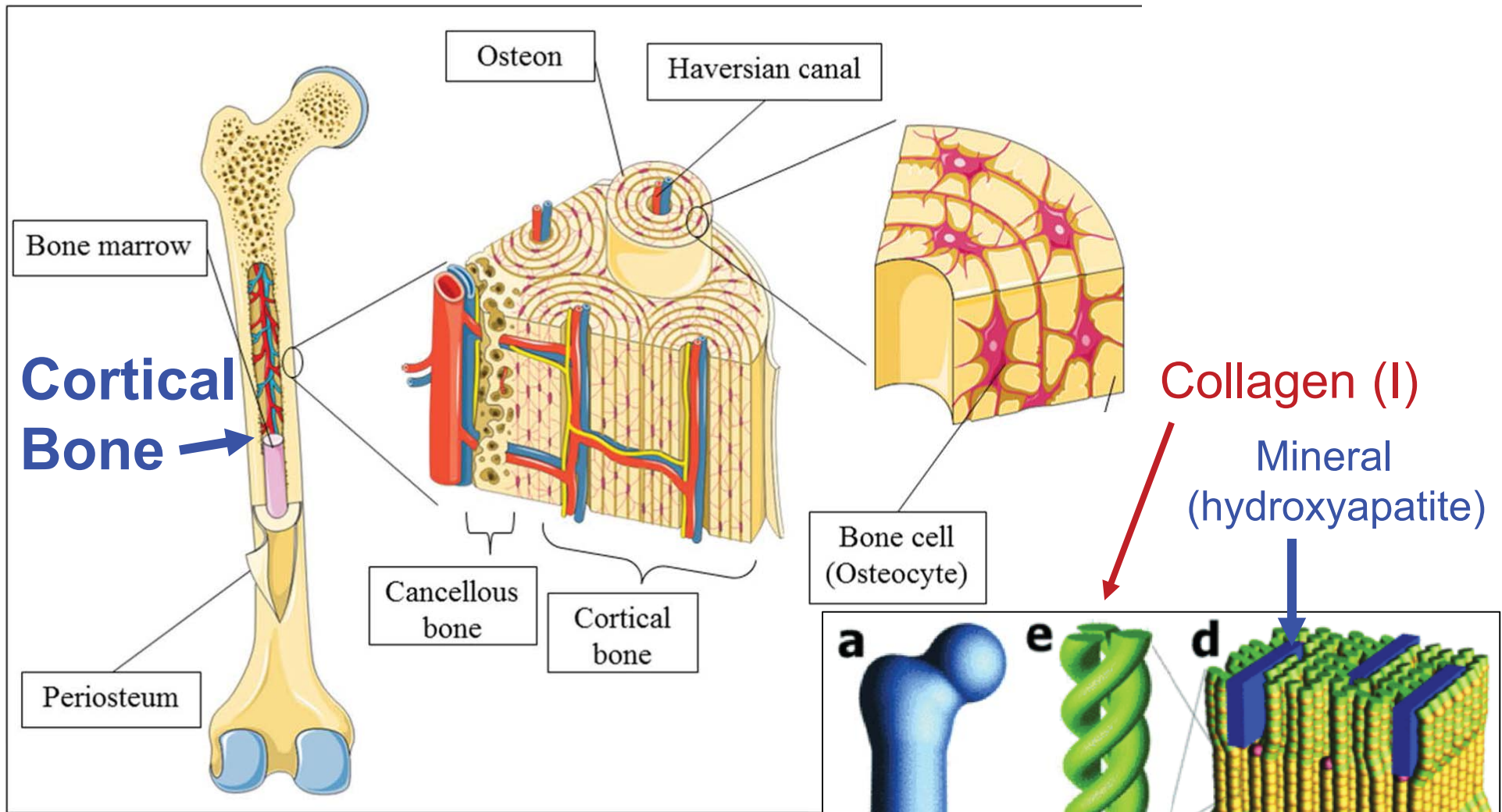
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Courtesy of Elsevier, Inc., <http://www.sciencedirect.com>. Used with permission. Source: Gutsman, Thomas. "Force Spectroscopy of Collagen Fibers to Investigate their Mechanical Properties and Structural Organization." *Biophysical Journal* 86, no. 5 (2004): 3186-93.

Stress vs strain curve of a rat tail tendon: (A-B) Toe - heel region, (C) linear region, (D) plateau, (E) rupture of the tendon.

(Gutsman+, Biophys J, 2004)

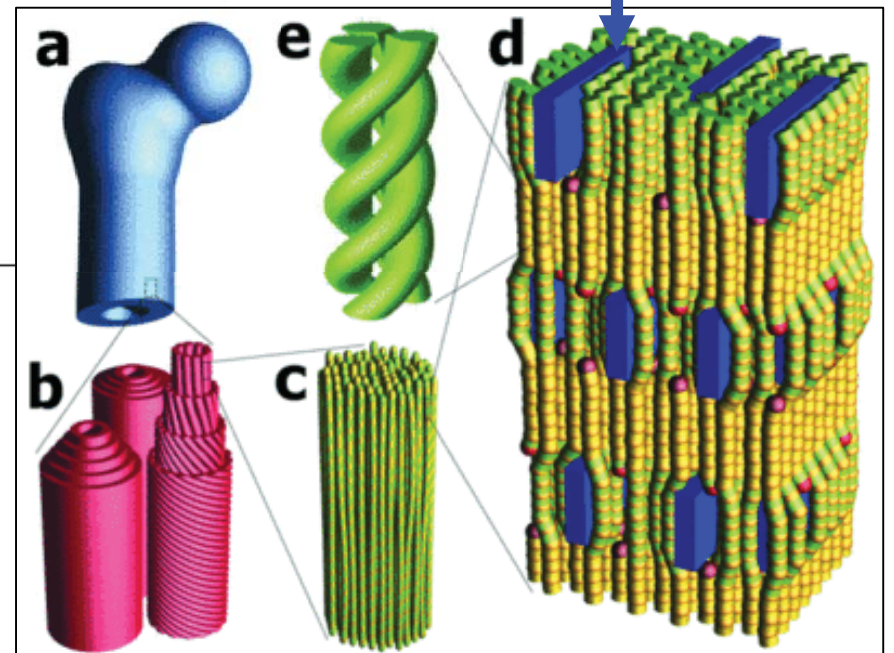


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Source: Bao, Chao Le Meng, et al. "Advances in Bone Tissue Engineering."

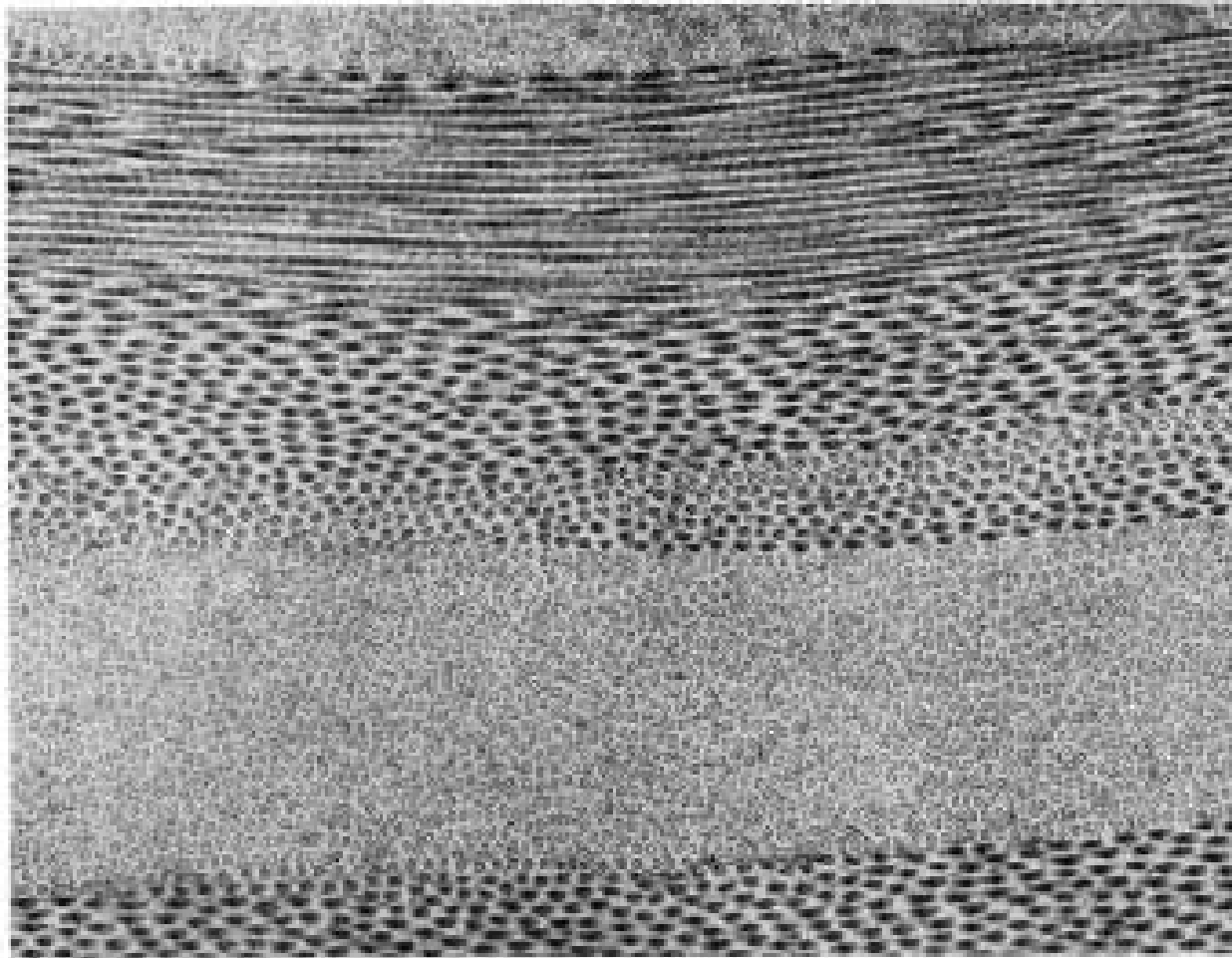
FY[YbYfUhj] Y A YX]V[bY UbX H]ggi Y '9b[]bYYf]b[(2013): 599-614.

Bone



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 Source: Zhou, Hongwen, et al. "Small-angle X-ray Study of the Three-dimensional Collagen /Mineral Superstructure in Intramuscular Fish Bone." *5dd]YX'7fmgU'c[fUd\m(2007).*

Corneal Stroma (normal)



~2 μm

Poisson's Ratio $\nu \sim 0$ HOW?

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(Text book page 241)

Regulation of gene expression in intervertebral disc cells by low and high hydrostatic pressure

(Eur Spine J, 2006)

Cornelia Neidlinger-Wilke · Karin Würtz ·
Jill P. G. Urban · Wolfgang Börm · Markus Arand ·
Anita Ignatius · Hans-Joachim Wilke ·
Lutz E. Claes

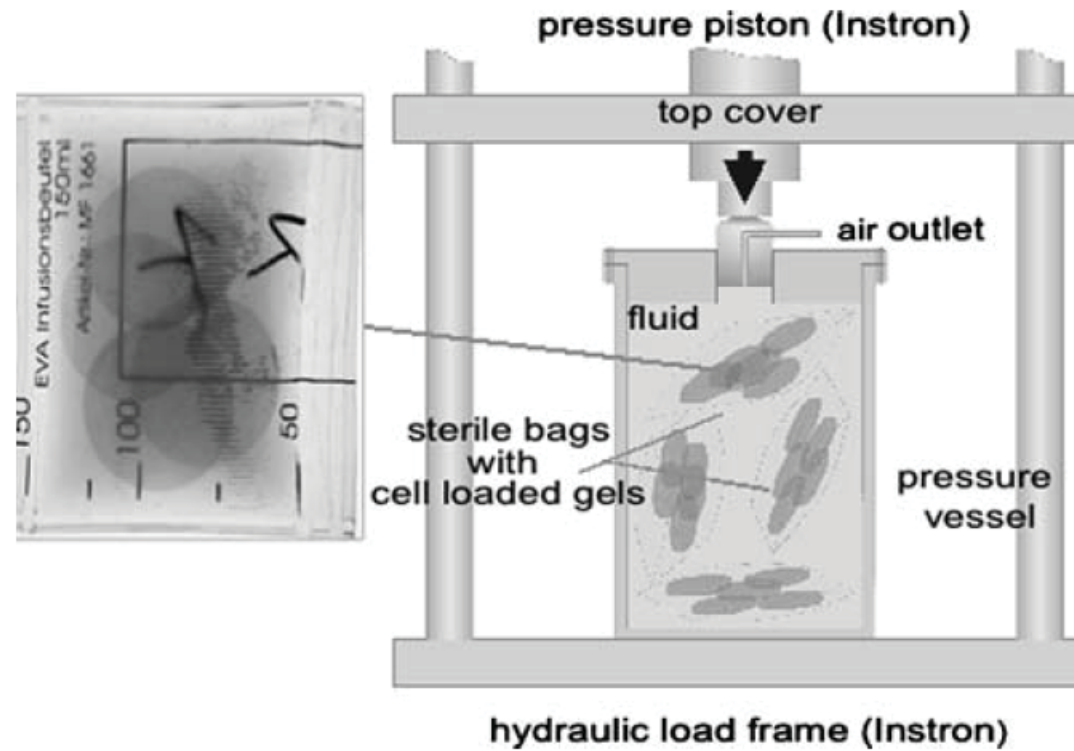


Fig. 1 Scheme of the stimulation device for the application of high hydrostatic pressure (2.5 MPa) and photo of a sterile bag with cell-seeded collagen gels

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Source: Neidlinger-Wilke, Cornelia, et al. "Regulation of Gene Expression in Intervertebral Disc Cells by Low and High Hydrostatic Pressure." *Eur Spine J* 15, no. 3 (2006): 372-8.

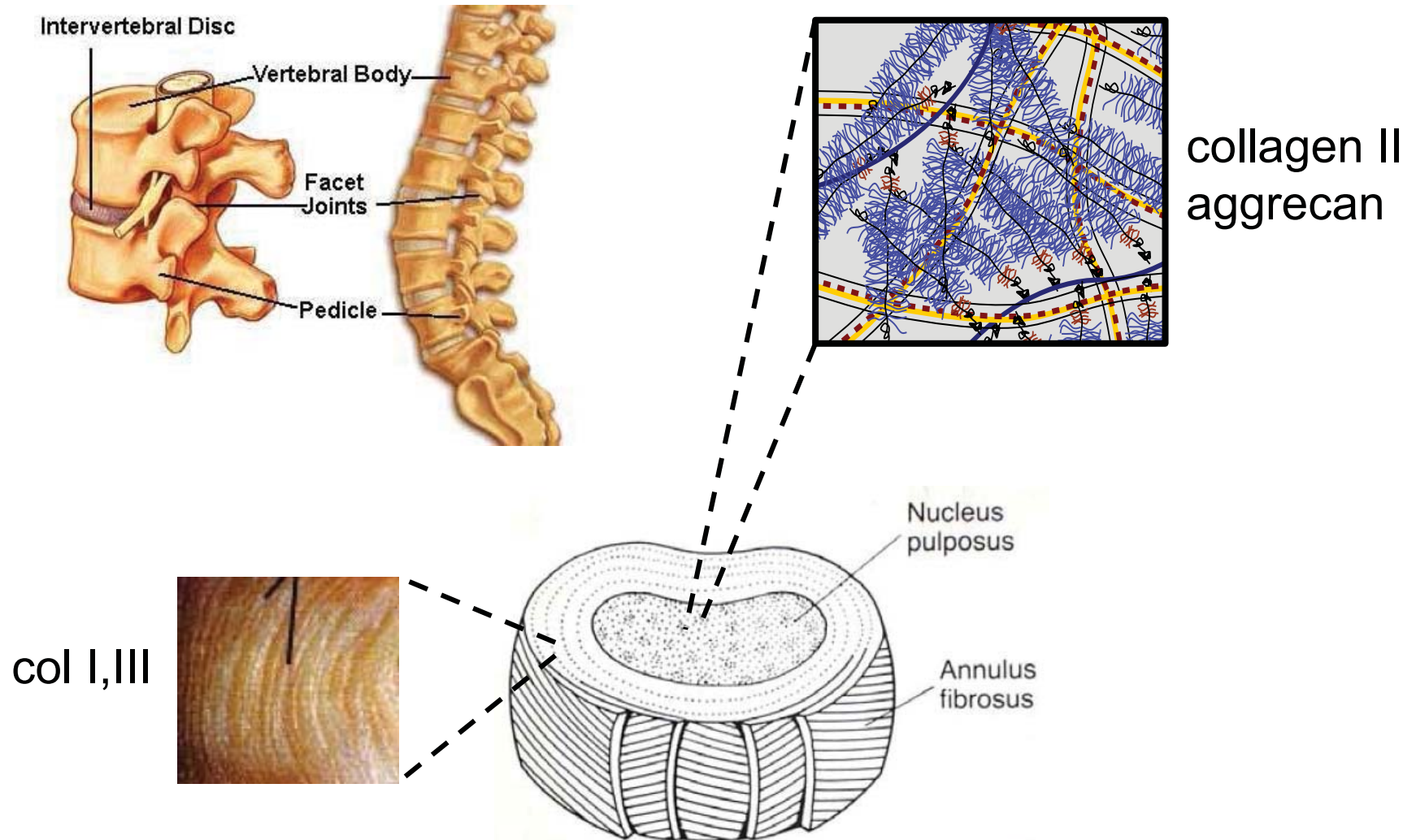
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(λ, G)	$\lambda + \frac{2G}{3}$	$\frac{G(3\lambda+2G)}{\lambda+G}$	λ	G	$\frac{\lambda}{2(\lambda+G)}$	$\lambda + 2G$

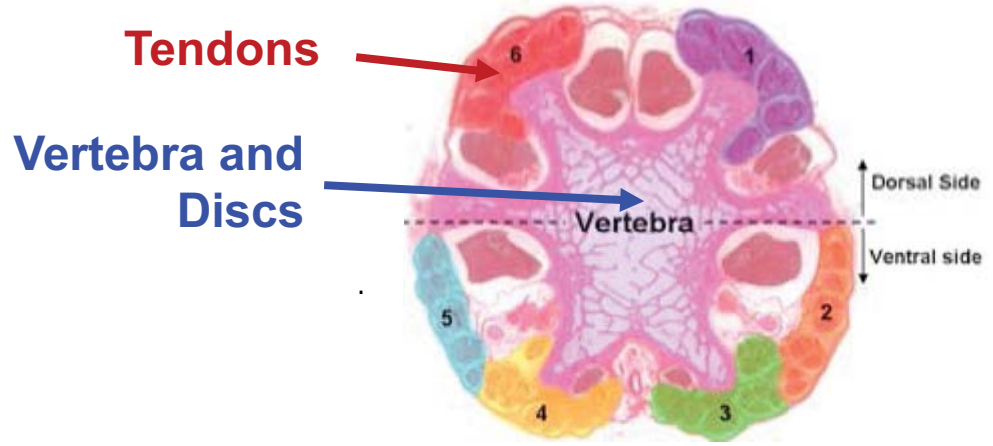
Intervertebral Disc

(Peter Roughley, Spine, 2004)

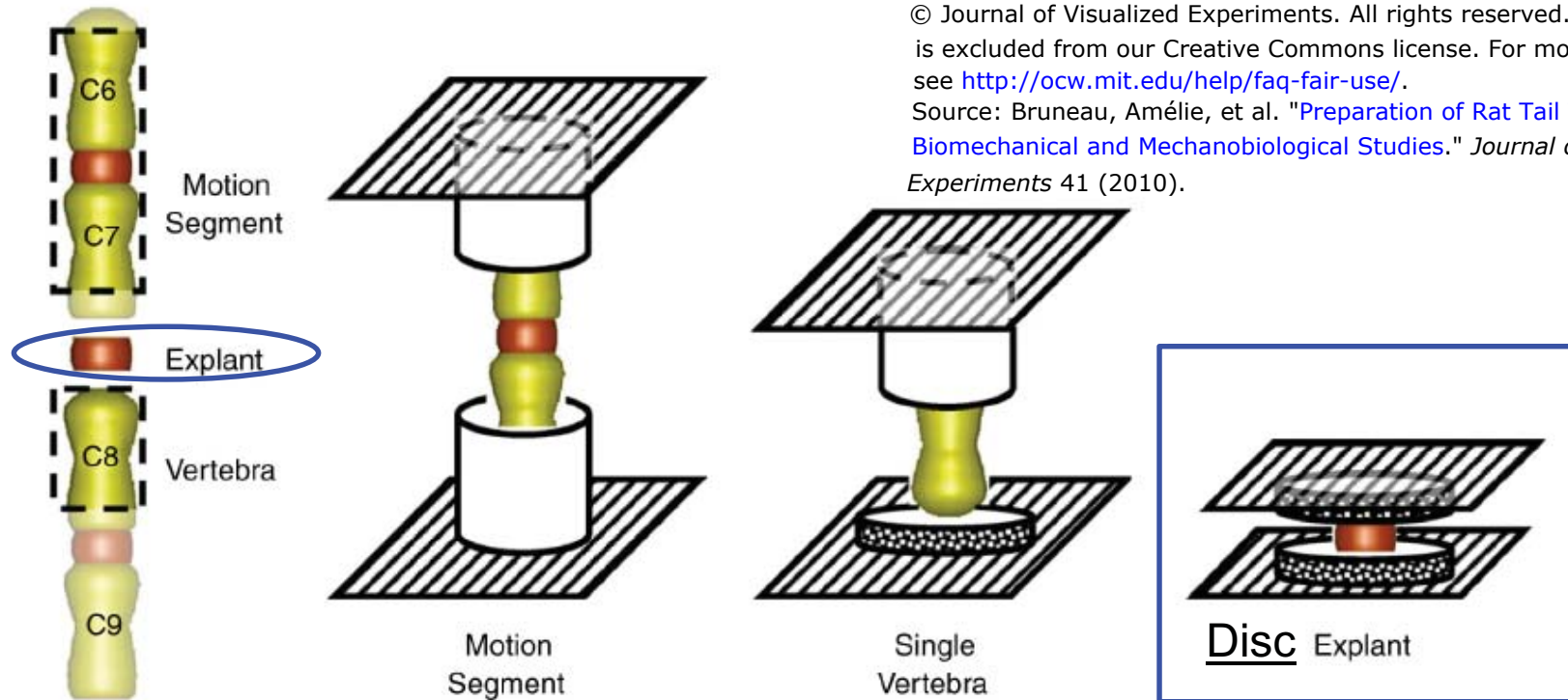


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Source: Roughley, Peter J. "Biology of Intervertebral Disc Aging and Degeneration: Involvement of the Extracellular Matrix." *GdjbY* 29, no. 23 (2004): 2691-9.

“Creep-Compression” of intervertebral disc (rat tail)



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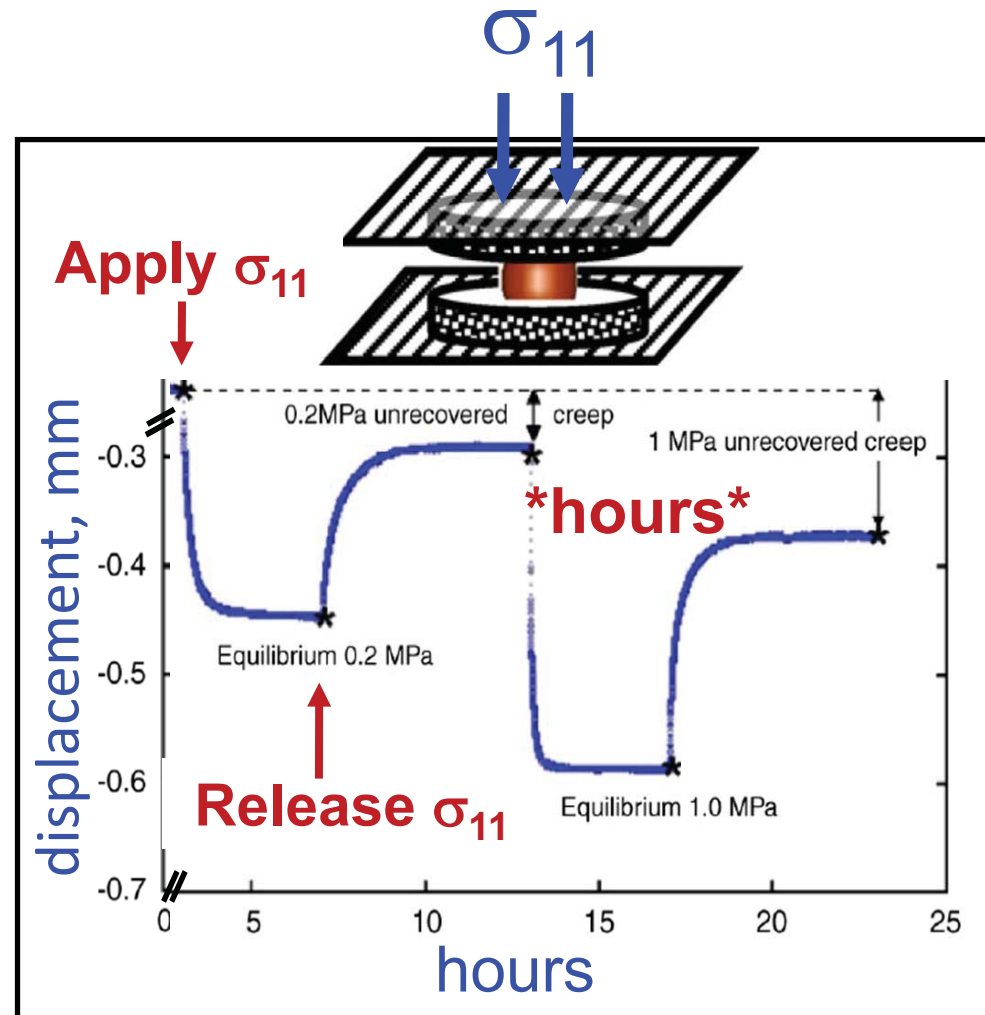


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(MacLean+, J Biomechanics, 2007)

“Creep-Compression” of intervertebral disc (rat tail):

Apply constant stress (σ_{11}) and measure displacement (strain) vs time



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Source: MacLean, Jeffrey J. "Role of Endplates in Contributing to Compression Behaviors of Motion Segments and Intervertebral Discs." -*RXUDDORI %LRP HFKDQLFV* 40, no. 1 (2007): 55-63.

(MacLean+, J Biomechanics, 2007)

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